

## Practical Synthesis of Enantiomerically Pure *myo*-Inositol Derivatives

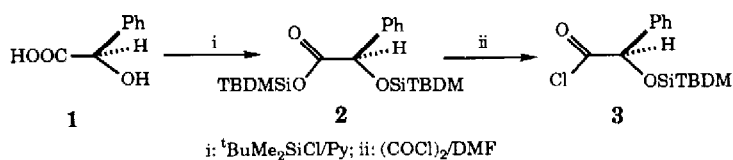
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**Abstract:** The synthesis of enantiomerically pure *myo*-inositol derivatives is accomplished using a mandelic acid-derived acyl protecting group.

A number of synthetic schemes leading to enantiomerically pure, and properly functionalized derivatives of *myo*-inositol start from one of the three positional isomers of di-*O*-cyclohexylidene-*myo*-inositol.<sup>1,2</sup> These derivatives are formed as a mixture, and thus have to be separated by a combination of crystallization and chromatographic techniques,<sup>2</sup> making their availability difficult. In contrast, *cis*-monoacetals of *myo*-inositol are synthesized in one step in good yield.<sup>3</sup> Recently we have found that *cis*-monoacetals can be selectively protected at the 1-hydroxyl group with bulky acylating or silylating reagents.<sup>4</sup> By applying selective protection of the remaining hydroxyl groups a number of useful intermediates for the synthesis of phosphoinositides can be obtained. In this communication we report on the synthesis of novel synthetic precursors of inositol-1,4,5-trisphosphate (IP<sub>3</sub>), phosphatidylinositol (PI), and phosphatidylinositol-4,5-bisphosphate (PIP<sub>2</sub>). Our approach is to combine the selective protection of 1-hydroxyl group and formation of diastereomeric esters with a chiral auxiliary, and hence to eliminate the need for additional steps to form separable diastereomeric esters of inositol.

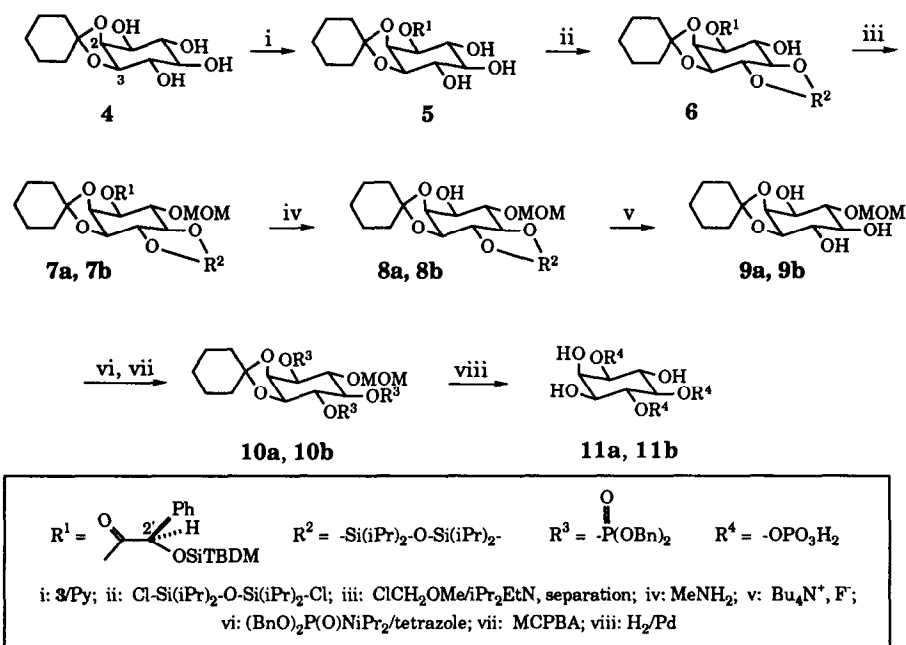
Silylation of (R)-(-)-mandelic acid (**1**) with 2 equiv. of *tert*-butyldimethylsilyl chloride (TBDMS-Cl) in pyridine and subsequent chlorination of the resulting silyl ester **2** with oxalyl chloride/DMF<sup>5</sup> afforded 2-(*tert*-butyldimethylsilyloxy)phenylacetyl chloride (**3**).<sup>6</sup>



Reaction of cyclohexylidene-*myo*-inositol (**4**) with **3** in pyridine at -40°C afforded the mixture of 1-protected diastereomeric esters **5** (55%). Formation of other positional isomers could not be detected. The diastereomers **5** proved to be difficult to separate by chromatographic methods.<sup>7a</sup> However, simple precipitation from ether-hexane afforded one of the isomers **5a** with >96% d.e.<sup>7b</sup> The mixture of diastereomers remaining in the mother liquor was chromatographed to remove higher protected

derivatives, and was further treated with 1,3-dichloro-1,1,3,3-diisopropylidisiloxane in pyridine at room temperature. The resulting diastereomeric mixture of 6-alcohols (**6a** and **6b**) was reacted with methoxymethylene chloride - diisopropylethylamine in THF at 55°C during 12 h (62%). Fully protected compounds **7a,b** were easily separated into individual isomers by chromatography on silica gel (hexane-ether, 40:1, **7a**:  $R_F$  0.18; **7b**:  $R_F$  0.13).<sup>8</sup> The isomers of **7** were converted into functionally useful enantiomeric alcohols **8**<sup>9</sup> by deacylation at the 1-position with methylamine (83%), and subsequently into triols **9**<sup>10</sup> by desilylation of **8** with tetra-*n*-butylammonium fluoride (100%).

The combination of acyl, silyl and acetal protective groups in **5-9** make these compounds versatile starting materials for the synthesis of phosphatidylinositol, inositol-1-phosphate (from **5**<sup>11</sup> and **8**), 6-glycosylated phosphatidylinositols (from **6**), and PIP<sub>2</sub> (from **8**), in addition to the synthesis of IP<sub>3</sub> described below. The synthesis of various phosphorothioate analogs, and unsaturated fatty acid-containing inositol phospholipids should also be possible.



The utility of synthesized compounds as phosphoinositides precursors was demonstrated by the synthesis of L- and D-IP<sub>3</sub> starting from enantiomers **9a** and **9b**, respectively.<sup>12</sup> This synthesis also enabled determination of the inositol configurations in **5-9** on the basis of configurations of IP<sub>3</sub>. Thus, triols **9** were treated with *N,N*-diisopropyl-*O,O*-dibenzylphosphoramidite/tetrazole, and the resulting trisphosphites were oxidized with *m*-chloroperbenzoic acid to give the corresponding tris(dibenzyl-

phosphates) **10a,b**. Deprotection of the phosphate groups in **10** with  $H_2/Pd$  in methanol and subsequent spontaneous cleavage of inositol protecting groups during hydrogenolysis afforded  $IP_3$  (**11**). The synthesis starting from the crystalline triol **5a** produced enantiomerically pure L-*myo*-inositol-1,4,5-trisphosphate (**11a**). D- $IP_3$  (**11b**) was synthesized in a reaction sequence starting from the more polar isomer **7b**.

The advantage of our method is in its brevity, versatility of intermediates and in a fewer number of chromatographic purifications required along the synthesis of phosphoinositides precursors.

*This project has been supported by a Grant GM 30327 from National Institutes of Health*

#### References and Notes:

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- Reagent **3** was sufficiently pure in a crude form;  $^1H$  NMR ( $CDCl_3$ )  $\delta$  7.6-7.45, (m, Ph, 5H), 5.52 (s, H-2, 1H), 1.04 (s, Bu, 9H), 0.14, 0.05 (each s, Me, 3H).
- (a) Chromatographic separation was attempted for triols **5**, and the corresponding exhaustively acetylated and methoxymethylated derivatives. (b) **5a**:  $^1H$  NMR ( $CD_3OD$ )  $\delta$  7.5-7.3 (m, Ph, 5H), 5.41 (s, H-2', 1H), 4.93 (dd, H-1, J 4.3, 9.8 Hz, 1H), 4.26 (dd, H-2, J 4.4, 5.0 Hz, 1H), 3.91 (dd, H-3, J 5.2, 7.5 Hz, 1H), 3.75 (tr, H-6, J 9.6 Hz, 1H), 3.47 (dd, H-4, J 7.5, 9.9 Hz, 1H), 3.19 (dd, H-5, J 9.9, 9.6 Hz, 1H), 1.6-1.2 (m,  $CH_2$ , 10H), 0.92 (s, Bu, 9H), 0.14, 0.05 (s, Me, 3H).
- 7a**:  $^1H$  NMR ( $CDCl_3$ )  $\delta$  7.45, 7.3 (m, Ph, 5H), 5.24 (s, H-2', 1H), 4.98 (dd, H-1, J 3.9, 10.2 Hz, 1H), 4.51, 4.45 (d,  $CH_2$ , 2H), 4.42 (dd, H-2, J 4.0, 5.3 Hz, 1H), 3.98 (dd, H-3, J 5.3, 6.8 Hz, 1H), 3.84 (tr, H-6, J 9.0 Hz, 1H), 3.76 (dd, H-4, J 6.8, 9.4 Hz, 1H), 3.52 (tr, H-5, 9.2 Hz, 1H), 3.12 (s, Me, 3H), 1.7-1.0 (m,  $CH_2$ , iPr, 38H), 0.89 (s, Bu, 9H), 0.10, 0.02 (s, Me, 6H); Compound **7a** had the same inositol configuration as triol **5a**. **7b**: 7.45, 7.3 (m, Ph, 5H), 5.32 (s, H-2', 1H), 4.96 (dd, H-1, J 4.0, 10.0 Hz, 1H), 4.91, 4.71 (d,  $CH_2$ , 2H), 4.28 (tr, H-2, J 4.2 Hz, 1H), 3.98 (tr, H-6, J 9.2 Hz, 1H), 3.91 (dd, H-3, J 5.0, 6.8 Hz, 1H), 3.72 (dd, H-4, J 6.8, 9.3 Hz, 1H), 3.53 (tr, H-5, 9.2 Hz, 1H), 1.6-1.0 (m,  $CH_2$ , iPr, 38H), 0.92 (s, Bu, 9H), 0.13, 0.02 (each s, Me, 3H)
- 8a**:  $^1H$  NMR ( $CDCl_3$ )  $\delta$  4.86, 4.76 (each d,  $CH_2$ , 2H), 4.43 (dd, 1H, H-2, J 3.5, 5.1 Hz), 3.99 (dd, H-1, J 5.2, 6.8 Hz, 1H), 3.74 (m, 3H), 3.50 (tr, H-5, J 6.8 Hz, 1H), 3.45 (s, Me, 3H), 1.6 (br m), 1.40 (br m), 1.25 (m, 2H), 1.05 (m, 38H);  $^{13}C$  NMR ( $CDCl_3$ )  $\delta$  110.4 ( $C_q$ ), 98.3 ( $OCH_2$ ), 80.9, 79.6, 79.3, 77.1, 75.1, 69.3 (CH-inositol), 55.7 (OMe), 37.9, 35.2, 24.9, 23.9 (CH-isopropyl), 17.3-16.9 (5 peaks,  $CH_2$ ), 12.7, 12.6,

- 12.8, 11.8 (Me);  $[\alpha]_{\text{D}}^{20} +7.5^\circ$  (c 4.2,  $\text{CHCl}_3$ ); **8b**:  $[\alpha]_{\text{D}}^{20} -7.9^\circ$  (c 4.2,  $\text{CHCl}_3$ ).
10. **9a**:  $[\alpha]_{\text{D}}^{20} -20.5^\circ$  (c 4.4, MeOH);  $^1\text{H}$  NMR ( $\text{CD}_3\text{OD}$ )  $\delta$  4.85 (m,  $\text{CH}_2\text{O}$ ), 4.35 (dd, H-2, J 4.0, 5.2 Hz, 1H), 3.93 (dd, H-1, J 5.2, 7.5 Hz, 1H), 3.78 (dd, H-3, J 4.0, 9.2 Hz, 1H), 3.61 (tr, H-4, J 9.0 Hz, 1H), 3.58 (dd, H-6, J 7.5, 9.9 Hz, 1H), 3.45 (s, Me, 3H), 3.22 (dd, H-5, J 9.0, 9.9 Hz, 1H), 1.8-1.55 (m,  $\text{CH}_2$ , 10H); **9b**:  $[\alpha]_{\text{D}}^{20} +15.5^\circ$  (c 2.5, MeOH).
11. Exhaustive methoxymethylation of **5a** followed by deacylation produces enantiomerically pure 1-alcohol suitable for phosphorylation.
12. **11a** (from **7a**),  $[\alpha]_{\text{D}}^{20} +31.3^\circ$  (c 2.6,  $\text{H}_2\text{O}$ , as pentahydrate of hexasodium salt, calculated for acid form), lit.<sup>13</sup>  $+35^\circ$ ;  $^{31}\text{P}$  NMR ( $\text{D}_2\text{O}$ , acid form)  $\delta$  -0.6, -1.0, -1.7 ppm;  $^1\text{H}$  NMR ( $\text{D}_2\text{O}$ , acid form)  $\delta$  4.23 (dtr, H-4, J 9.1 Hz, 1H), 4.15 (tr, H-2, J 2.8 Hz, 1H), 3.96 (m, H-1, H-5, 2H), 3.76 (tr, H-6, J 9.5 Hz, 1H), 3.59 (dd, H-3, J 2.7, 9.8 Hz, 1H); **11b**:  $[\alpha]_{\text{D}}^{20} -30.5^\circ$  (c 1.3,  $\text{H}_2\text{O}$ ), lit.<sup>13</sup>  $-30^\circ$ ;  $^1\text{H}$  and  $^{31}\text{P}$  NMR spectra were essentially the same as those of **11a**.
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(Received in USA 28 October 1991)